SureScreen Scientifics

LAB SAMPLE ID
Internal Use Only

Filtration eDNA Sample Form

*Required Fields

Contact

Name*:	
Company*:	Phone*:
Report Email*:	

Payment

Invoice Email*:	☐ Same as report email
Invoice Address*:	
PO/Reference:	Note: Analysis will NOT be started unless you have provided us with a PO/Reference **or** ticked the Pay By Card option. For samples in batches, each unique PO/Reference will get its own report and invoice. **For new customers, payment is required before results are reported.**
New Customer/ Pay by Card: ☐ If you are a new customer **OR** wish to pay by card please tick this box.	

Service

Turnaround time*:

☐ 10 working days

☐ 5 working days

***Note** For a detailed breakdown of pricing, please refer to the Target Species section. All prices are per sample. If no box is ticked, we will assume a 10 working day service is required.

Due to the nature of print, these prices are subject to change. Please refer to our website for the latest information.

Target species*:	10 working days	5 working days
☐ One target species	£150 +VAT	£250 +VAT
☐ Two target species	£185 +VAT	£285 +VAT
☐ Three target species	£220 +VAT	£320 +VAT
☐ Four target species	£255 +VAT	£355 +VAT

Sample Info

Unique Sample Name/ID*:	
Site*:	Date*:
O\S Reference*:	Volume Filtered*:

Pick up to four target species

☐ All UK Shad	☐ Brown trout	☐ Crucian carp	☐ Natterjack toad	☐ Smooth newt
☐ Alpine newt	☐ Chinese mitten crab	☐ European eel	☐ Pike	☐ Spined loach
☐ American mink	☐ Chytrid fungus *Bd*	☐ European perch	☐ Quagga mussel	☐ Spiny-cheek crayfish
☐ Arctic charr	☐ Chytrid fungus *B.sal*	☐ European smelt	☐ Rainbow trout	☐ Topmouth gudgeon
☐ Asian clam	☐ Common carp	☐ Freshwater pearl	☐ Red-swamp crayfish	☐ Water vole
☐ Atlantic salmon	☐ Common frog	☐ Great crested newt	☐ Rudd	☐ White-clawed crayfish
☐ Blue-green algae	☐ Common toad	☐ Marbled crayfish	☐ Sea lamprey	☐ Zebra mussel
☐ Brook/River lamprey	☐ Crayfish plague	☐ Narrow-clawed crayfish	☐ Signal crayfish	☐ Other (specify below)

Other:

If your target species is not in our list, please check with our team before sending your samples in.

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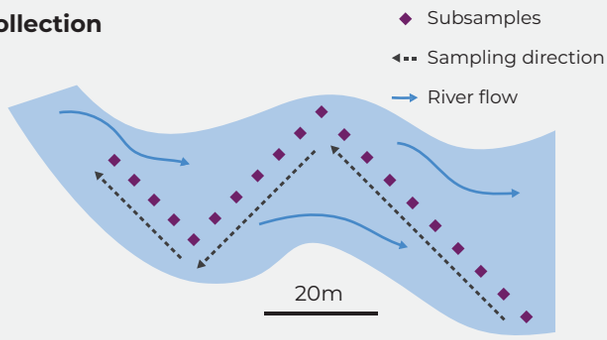
Filtration eDNA Sample Form

Instructions

Instructions for sample collection



1. Identify 20 sites around the pond/river where you plan to collect your subsamples from. These should be spaced as evenly as possible around the site. In rivers, samples should be taken in an upstream diagonal pattern where possible, if it is necessary to enter the watercourse. Alternatively, you can collect samples along the perimeter of a pond or along both shores of a river, using a telescopic pole to obtain subsamples from areas difficult to access or which are further from the river bank.
2. Put on the gloves provided and open the bag.
3. Using the 30ml ladle provided, collect a subsample from at least 5-10cm deep from each of the sites previously identified in step 1 (total 20 subsamples). The water sample should be taken from the middle of the water column. Where possible, avoid any disruption of sediment as this can both clog the filter quicker and introduce ancient DNA into the sample. Transfer each ladle full of water to the bag provided In larger sites it may be necessary to use a telescopic pole.
4. Once all sites have been sampled, tightly scrunch the bag and shake vigorously for 10 seconds (to mix any DNA within the sample equally).
5. Using the large syringe, take 50ml of sample and attach the syringe using a half twist action to the narrow end of the filter unit (the syringe will only fit to one end of the filter). Apply pressure to the syringe until all liquid has passed into and through the filter unit. Note, twistng too far can damage the luer lock connection on the filter. Remove the filter unit from the syringe and repeat this step until up to 500ml (minimum required volume = 150ml) is filtered/the filter becomes clogged/you are no longer able to push any liquid through. The more liquid passed through the filter unit, the more reliable results will be, however, be careful not to exert too much force as the filter casing can crack under extreme pressure. If/when resistance becomes too high, finish filtering the sample. Record the amount of liquid which has been filtered on this sheet.
6. Empty the syringe and fill with air, attach this to the filter and repeatedly push air through the filter until it is free of water
7. Screw one white cap onto the thick end of the filter unit. Place to one side.
8. Carefully take the white cap from the small pre-filled blue syringe, this contains an excess of the preservative solution. Place the white cap to one side, connect the syringe to the open end of the filter unit and apply gentle pressure until all 2ml of solution is stored within the filter casing.
9. Screw the white cap from step 8 to the narrow end of the filter, ensure both cap ends are tight, and then place the filter into the 50ml storage tube provided.
10. Finally, fill in the sample collection form (on the reverse of this page).
11. Place the 50ml tube containing the sealed filter and the large syringe (this helps us reduce plastic waste in the lab) in the clear plastic bag and return to the laboratory address below for analysis, with the corresponding analysis form.
12. Results will be emailed to you within the specified turnaround time.

Detailed sample collection guidance
For further assistance with sample collection, visit our website or scan this QR code to access our detailed step-by-step filtration sample collection guide.

Tips

- Kit components are single use only and must not be reused for other samples.
- If storage of samples is necessary before returning to the lab, samples should be refrigerated where possible.
- At a maximum, preservative filled samples can be kept at room temp for 2 weeks prior to analysis, longer if chilled.
- Sending in a batch of samples? No need to fill out contact details multiple times, just include it on one of the forms in the box and we will work out the rest!
- **Help us save on single-use plastics in the analysis of your sample by returning the syringes with the kit**
- We can now recycle plastic kit components; please send back gloves and ladles for responsible recycling

Returns your kits to:
SureScreen Scientifics, Morley Retreat, Church Lane, Morley, Derbyshire, DE7 6DE

Have you used our other services?
www.SureScreenScientifics.com