

**LAB SAMPLE ID**

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**Note: Please include a PO or Ref No. for this work, analysis will NOT be started until a valid PO has been received.**

For samples in batches, each unique PO will have its own report and invoice. Report can be sent to multiple emails if required. For new customers, payment is required before results are reported.

**PURCHASE ORDER\*:**

Please write any important notes here:

**Filtration eDNA Sample Form**

**ORDER**

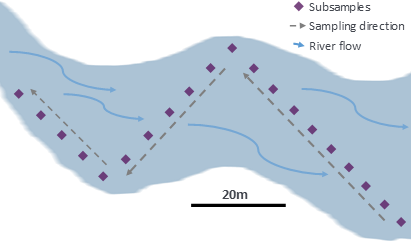
**NOTES**

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| **CONTACT** | **\*Required Fields**  NAME\*: Type here… |
| COMPANY\*: |
| EMAIL\*: |
| TELEPHONE\*: |
| INVOICE ADDRESS\*:  *See our website for current prices.* |

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| **SAMPLE INFO** | SAMPLE NAME/ID\*:  This will be the unique sample identifier on your report |
| SITE NAME\*: DATE: |
| O/S REFERENCE\*: VOLUME FILTERED\*: |
| **PLEASE SELECT SPECIES FOR ANALYSIS: (up to a maximum of 4 can be analysed per kit):**      Atlantic salmon  Brown/sea Trout  Common carp  Crucian carp  Pike  Chytrid (*B. dendrobatidis*)  Chytrid (*B. salamandrivorans*)  Asian clam  Sea lamprey  Other (please specify below)  Freshwater pearl mussel  Zebra mussel  Quagga mussel  Great crested newt  Smooth newt  White-clawed crayfish  Signal crayfish  Marbled crayfish  Crayfish plague  European eel                                        Other:    *If your target species is not in our list, please check with our team before sending your samples in!* |

This form is also available to download and edit as a word document at: **SureScreenscientifics.com/forensic-ecology/other-species**

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| **INSTRUCTIONS** | **INSTRUCTIONS FOR SAMPLE COLLECTION**   1. Identify 20 sites around the pond/river where you plan to collect your subsamples from. These should be spaced as evenly as possible around the site. In rivers, samples should be taken in an upstream diagonal pattern where possible, if it is necessary to enter the watercourse.   Alternatively you can collect samples along the perimeter of a pond or along both shores of a river, using a telescopic pole to obtain subsamples from areas difficult to access or which are further from the river bank.   1. Put on the gloves provided and open the Whirl-Pak bag by tearing the strip at the perforated line. Gloves should be worn at all times during collection. 2. Using the 30ml ladle provided, collect a subsample from at least 5-10cm deep from each of the sites previously   identified in step 1 (total 20 subsamples). The water sample should be taken from the middle of the water column. Where possible, avoid any disruption of sediment as this can both clog the filter quicker and introduce ancient DNA into the sample. Transfer each ladle full of water to the Whirl-pak bag provided In larger sites it may be necessary to use a telescopic pole.  Once collected close the bag securely and shake to mix the water sample.   1. Once all sites have been sampled, fold the top of the bag over and shake the Whirl-Pak bag vigorously for 10 seconds (to mix any DNA within the sample equally). 2. Using the large syringe, take 50ml of sample and attach the syringe using a half twist action to the narrow end of the filter unit (the syringe will only fit to one end of the filter). Apply pressure to the syringe until all liquid has passed into and through the filter unit. Note, twisting too far can damage the luer lock connection on the filter. Remove the filter unit from the syringe and repeat this step until up to 500ml (minimum required volume = 150ml) is filtered/the filter becomes clogged/you are no longer able to push any liquid through. The more liquid passed through the filter unit, the more reliable results will be,   however, be careful not to exert too much force as the filter casing can crack under extreme pressure. If/when resistance becomes too high, finish filtering the sample. Record the amount of liquid which has been filtered on this sheet.   1. Empty the syringe and fill with air, attach this to the filter and repeatedly push air through the filter until it is free of water. 2. Screw one red cap onto the thick end of the filter unit. Place to one side. 3. Carefully take the red cap from the small pre-filled blue syringe, this contains an excess of the preservative solution.   Place the red cap to one side, connect the syringe to the open end of the filter unit and apply gentle pressure until all 2ml of solution is stored within the filter casing.   1. Screw the red cap from step 8 to the narrow end of the filter, ensure both cap ends are tight, and then place the filter into the 50ml storage tube provided. 2. Finally, fill in the sample collection form (on the reverse of this page) 3. Place the 50ml tube containing the sealed filter and the large syringe (this helps us reduce plastic waste in the lab) in the clear plastic bag and return to the laboratory address below for analysis, with the corresponding analysis form. 4. Results will be emailed to you within 10 working days of sample receipt.   **Detailed sample collection guidance**  For further assistance with sample collection, visit our website or scan this QR code to access our detailed step-by-step filtration sample collection photo-guide and FAQ’s. |
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| **TIPS** | * Kit components are single use only and must not be reused for other samples. * If storage of samples is necessary before returning to the lab, samples should be refrigerated where possible.   At a maximum, preservative filled samples can be kept at room temp for 2 weeks prior to analysis, longer if chilled.   * Sending in a batch of samples? No need to fill out contact details multiple times, just include it on one of the forms in the box and we will work out the rest! * **Help us save on single-use plastics in the analysis of your sample by returning the syringes with the kit**   Any questions? Email us at [**eDNA@SureScreen.com**](mailto:eDNA@SureScreen.com) |

**RETURN YOUR KITS TO:**

SureScreen Scientifics, Morley Retreat, Church Lane, Morley, Derbyshire, DE7 6DE

**Have you used our other services?**

# [www.SureScreenScientifics.com/Forensic-Ecology](http://www.SureScreenScientifics.com/Forensic-Ecology)