**\*NAME:** …………………………………………………………..………………. **\*COMPANY:** ……………………….……………………………………………

**\*EMAIL:** …..…………………………………………………………………..……………………………………………………………………………………………….

**\*INVOICE ADDRESS:** ………………………………………………………………………………………………………………………………………………………

**\*TEL:** ………………………………………………………………………. **\*PURCHASE ORDER** ……………………………………………………………………

*Note: Please include a PO for this work, analysis will NOT be started until a valid PO has been received. When samples are sent in batches, each individual PO will be its own report and invoice. The report can be sent to multiple emails if required.*

**Crayfish eDNA - Sample Collection Form**

1. Identify where 20 sub-samples will be taken from the pond/river. The location of these should be spaced as evenly as possible around the site. In rivers, samples should be taken against the flow of the stream, working upstream in a diagonal pattern where possible to ensure that any disturbed ancient DNA is not collected and to avoid contamination, should it be necessary to enter the watercourse.

2. Open the sterile Whirl-Pak bag and collect 20 samples of water from around the site using the ladle. Once collected close the bag securely using the top tabs and shake for 10 seconds. This mixes DNA across the water sample.

3. Using the large sterile syringe, take 50ml of sample and then attach the syringe using a half twist action to the filter unit (the syringe will only fit to one end of the filter). Apply pressure to the syringe until all liquid has passed into and through the filter unit. Remove the filter unit from the syringe and repeat this step until 500ml is filtered/the filter becomes clogged/you are no longer able to push any liquid through. The more liquid passed through, the better. Record the amount of liquid which has been filtered on this sheet.

4. Empty the syringe and fill with air, attach this to the filter and push air through the syringe until it is completely free of water.

5. Screw one red cap onto the thick end of the filter unit. Place to one side.

6. Using the small syringe, collect 2ml of preservative solution, connect to the open end of the filter unit and apply gentle pressure until all 2ml of solution is stored within the filter casing. Repeat if necessary until filter casing is full of preservative.

7. Screw the red-caps to secure both ends of the filter and then place the filter into the 50ml tube provided. Sample can be stored in a cool dry place for a maximum of 3 weeks, longer if chilled, however for best results it is recommended to return to the laboratory as soon as possible.

**\*Required Fields**

**RELEVANT NOTES**

**LABORATORY SAMPLE ID**

Kit components are single use only and must not be reused for any other samples. Components can be returned to the laboratory for recycling. FAQ’s and a more detailed explanation as to how to collect a sample in different scenarios in available on our website.

**This form is available to download and edit as a word document at: www.surescreenscientifics.com/edna/crayfish**

**INSTRUCTIONS FOR SAMPLE COLLECTION**

*We currently only offer a standard turnaround time for this service of up to 10 working days.*

**\*SPECIES TO DETECT *(Please Tick)***

**White Clawed Crayfish**

**Signal Crayfish**

**Crayfish Plague**

***Note: There are different costs associated with analysing for one, two or three target species.***

**\*SAMPLE NAME**: ……………..………………………………………………………………….……..

**\*VOLUME OF WATER FILTERED:** ..….…………………………………………………………..

Site Name: ………………………………………………………………………………………………….

O/S Reference: …………………………………………………………………………………………… County: …………………………………… Sampling Date: ………………………………………..

SureScreen Scientifics

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**GREAT CRESTED NEWT (GCN) FORM**

